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Autoimmune Diseases in Limited Setting Country (Burkina Faso): Epidemiology, Technical and Diagnostic Performance of Specific Biomarkers for the Early Diagnosis

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Abstract

Background: Autoimmune diseases have been rarely reported in limited setting country. We aimed to investigate the burden, immunological and clinical diagnosis of autoimmune diseases in Burkina Faso.

Methods: We prospectively enrolled patients with a suspected autoimmune disease at University Hospital SOURO SANOU, in Bobo Dioulasso, BURKINA FASO, between 2012 and 2014 for this cohort study. Demographic characteristics, clinical manifestations, sera autoantibodies determination and plasma viral and bacteria status (HBV, HCV, HIV, and Dengue serotype 1, 2, 3 and 4, *Mycobacterium tuberculosis* and Lyme disease) were determined. All plasma samples were stored at - 80°C when analyzed was not making immediately.

Results: During the three year study period, totally 65 patients with suspicion of autoimmune diseases were diagnosed. The referral diagnosis was right in 17/65 patients (26.2%) among patients in the limited setting country (Burkina Faso). The incidence of autoimmune diseases was 17/73950 (0.023%). There were 4/73950 (0.005%) patients with systemic lupus erythematosus (SLE), 5/73950 (0.007%) with rheumatoid arthritis (RA), 2/73950 (0.003 %) with CREST and Polymyositis, 2/73950 (0.003%) with CREST, 1/73950 (0.001%) with Gougerot Sjögren's syndrome (SSG), 2/73950 (0.003%) with Thyroiditis of Hashimoto and, 1/73950 (0.001%) with sarcoidosis. The median age at the diagnosis of autoimmune diseases were respectively 51 years (28 -75 years) and male to female ratio of 0.06 (1/16). The plasma viral status (HBV, HCV, CMV and HIV, Lyme disease, Dengue serotype 1, 2, 3 and 4) was negative. *Mycobacterium tuberculosis*'s cases were 3.07%. The immune suppressor's treatment again autoimmune diseases were not efficient among 2/17 of patients.

Conclusion: The referral diagnosis was right in 17/65 (26.2%). The incidence was 17/73950 (0.023%) in the limited setting country (Burkina Faso). There was not a significant associated relation between autoimmune diseases and infection; there was not also significant association between autoimmune diseases and hygienic hypothesis.

Keywords: Autoimmune; Disease; Autoantibody

Abbreviations: ANA: Anti-nuclear Antibodies; CENP-B: Centromere protein B; dsDNA: Double-stranded DNA; ELISA: Enzyme-linked Immunosorbent Assay; IFI: Indirect Immunofluorescence Assay; Jo-1: Histidyl-t-RNA synthetase; AID: Autoimmune Disease; RNP70 A,C: Small nuclear ribonucleoprotein complexes; 70 kDa, A, C polypeptides Scl-70: DNA topoisomerase I; SLE: Systemic lupus erythematosus; SmD: Smith's antigen (a family of RNA-binding proteins); SSA/Ro: Sjögren's Syndrome A antigen/small ribonucleoprotein particle (Ro 52 and 60 kDa); SSB/La: Sjögren's Syndrome B antigen/Lupus antigen, La ribonucleoprotein domain family, member 3 ; U1RNP: U1 nuclear ribonucleoprotein (mixture of recombinant RNP70, A, C); RA: Rheumatoid Arthritis; HT: Hashimoto Thyroiditis

Introduction

Autoimmune diseases represent a heterogeneous family of chronic, disabling diseases with different natural histories and a wide spectrum of clinical symptoms. These disorders share underlying defects in the immune response leading the body to attack its own organs and tissues. Most of these diseases disproportionately affect women; however, persons of all racial, ethnic, and socioeconomic groups are affected [1,2]. Certain diseases, including systemic lupus erythematosus and scleroderma, are more common in African Americans, whereas others, such as type 1 diabetes and multiple sclerosis, are more common in Caucasians. All ages

are affected, with onset from childhood to late adulthood. While many individual autoimmune diseases are rare, collectively they are thought to affect approximately 5 to 8 percent of the United States population – 14 to 22 million persons [3-8]. Because of their chronicity, measured in decades, and their debilitating complications, autoimmune diseases exact high medical and socioeconomic costs. In addition, since autoimmune diseases affect women in their most productive years, their impact on families and society can be substantial. Although comprehensive national data on the incidence, prevalence, and medical and economic impact of autoimmune diseases do not exist in the aggregate, or for the majority of individual autoimmune diseases, the statistics that are available make clear

that the impact of these diseases is significant. Some of the available data for specific diseases are highlighted below. The purpose of our study is to establish immunology and clinical strategies for enhancing autoimmune diseases diagnosis in the limited setting country (BURKINA FASO) were incidence, prevalence, morbidity, and mortality of autoimmune diseases remains undiscovered.

Materials and Methods

Ethic Statement

Samples analyzed in our study derived from patients who went to hospital for consultation. Patients with autoimmune diseases enrolled in our study received effective immunosuppressors treatment alone or in association with another drug. The study was approved by our National Ethic Committee for health research in Burkina Faso Ouagadougou. Patients participating in this study gave written informed consent.

Patients and method

We prospectively enrolled patients who had a medical and immunology diagnosis of autoimmune diseases at University Hospital SOURO SANOU and another private clinical, in Bobo Dioulasso, BURKINA FASO, between January 2012 and November 2014 for this cohort study. Demographic characteristics, clinical manifestations, sera auto antibodies profile and plasma viral and bacterial statuses (HIV/HBV/HBC/CMV/ Dengue serotype 1, 2, 3 and 4 /Mycobacterium tuberculosis/Lyme disease) were determined.

In our study, autoimmune diseases diagnosis was a combination of autoantibody blood tests-rays, clinical presentation and blood tests that measure nutritional function and inflammation. In addition, a criterion of diagnosis was used to confirm each case of autoimmune disease without IFI method; the sera autoantibody profile of the patient represents an important prerequisite for the clinical diagnosis. Indeed, positive results for ANA and the presence of anti-double-stranded native DNA (anti-dsDNA) or anti-Sm antibodies constitute 2 of the 11 criteria for the diagnosis of systemic lupus erythematosus (SLE) [2,9-11]; positivity for ANA in high titer or the presence of anti-Ro/SSA or anti-La/SSB antibodies were diagnostic criteria for Sjögren's syndrome [10]; the presence of anti-Jo-1 antibodies was a criterion for the diagnosis of dermatopolymyositis [12,13]; the presence of anticentromere or anti-topoisomerase I (anti-Scl70) antibodies was the criterion for classifying subtypes of cutaneous systemic sclerosis as limited or diffuse [9].

Laboratory assay

The multiplex immunodot assay: We used for the diagnosis of autoimmune diseases the following Immunodot kit (biomedicals diagnostics, France)

- **ENA-DOT 7 kit** a qualitative enzyme immunoassay (EIA) for the simultaneous detection of 7 antibodies specificities: against Extractable Nuclear Antigens (SS-A, SS-B, Sm, Sm/RNP and Scl70), and against 2 additional antigens (centromere and Jo-1).
- **ANCA-MBG-DOT kit** a qualitative enzyme immunoassay (EIA) for the simultaneous detection of anti-neutrophil cytoplasm antibodies (ANCA) directed against Myeloperoxidase (MPO) and serine proteinase (PR 3). Anti-glomerular basement membrane (GBM) antibodies using a purified antigen based on type IV.
- **CYTO -DOT4 kit** a qualitative enzyme immunoassay (EIA) for the simultaneous detection of 4 auto-antibodies specificities against cytosol organists (anti-mitochondria, anti-Jo1, anti-ribosomes and anti -LK1)
- **DNA-DOT kit** a qualitative enzyme immunoassay (EIA) for the detection of anti-DNA antibodies directed against (DNA native).

- The antigens are coated on distinct membrane on the assay strip;
- First, the assay strip is incubated in a reaction tube containing the diluted sample of the patient. If this sample contains at least one of the antibodies, these antibodies will recognize and bind to the corresponding antigen. After incubation, a first wash step removes all of the unbound proteins.
- An alkaline phosphatase labelled protein A conjugate binds to the captured antibodies. The excess of unbound conjugate is removed by a second wash step.
- The bound conjugate is visualized with an insoluble substrate for the phosphatase enzyme (BCIP/NBT). This will give rise to a blue colored circle, confirming the presence of auto antibodies in the sample tested.
- **Anti-CCP antibodies** were measured using a two-step immunoassay whit the automated sample pretreatment for the semi quantitative determination of the Ig G class of auto antibodies specific to cyclic citrullinated peptide in human serum using chimiluminescent microparticle immunoassay (CMIA) technology whit flexible assay protocols referred to as chimiflex. Samples with results < 5 U/ml were defined as negative. Ig G and Ig M RF was determined using chimiluminescent microparticle immunoassay (CMIA) technology whit flexible assay protocols referred to as chimiflex. Samples with results < 20 UI/ml were defined as negative.

Screening for HIV, HBV and HCV: Chimiluminescent microparticle immunoassay (CMIA) technology (Architect *ci4100*[®]) was used in screening for HIV, HBV and HCV.

Screening for CMV and Lyme diseases: Enzyme linked Fluorescence Assay (ELFA) technology (MiniVidas[®]) was used in screening for CMV and Lyme diseases. Dengue serotype 1,2,3,4,5 and *Mycobacterium tuberculosis* were determined using the immunochromatographic technical (Hexagon D[®] and Hexagon TB[®] respectively).

Statistical analysis

Data were analyzed using Wilcoxon test to compare rates. We used Fischer (t) test for the percentages. Non parametric tests (Kruskal Wallis and Mann-Witney) were used when normality of distribution wasn't verified. Results were considered statistically significant when p<0.05. Analysis was performed with STATA[™].

Results

Patients and baseline characteristics

The number of visits during the study period was 73950 including 17 cases of autoimmune diseases. The referral diagnosis was right in 17/65 patients (26.2%). The incidence was 17/73950 (0.023%).

Table 1 shows demographic and clinical characteristics of autoimmune diseases patients in the study cohort. Comparing gender impact on the autoimmune diseases, the women were more likely to have autoimmune diseases than men (p=0.004). Analyzing correlation between autoimmune diseases and race or ethnicity and family annual incomes, no differences attributable to ethnicity and family annual incomes were observed (p=0.7).

Antibodies against HBV or HCV (Hepatitis B or C virus), HIV, CMV, *Borrelia burgdorferi* were found in sera or saliva of patients with autoimmune diseases such as SSG, SLE, and PR. No correlation was found in our study (p=0.3).

Inflammatory proteins profiles in the sera of patients with autoimmune diseases

The table 2 show inflammatory proteins profiles in the sera of patients with autoimmune diseases in the study cohort. Analyzing the different

Age (years)	
Median range	51 (28- 75)
Gender	
Male	4 (6.15%)
Female	61 (93.85%)
Family annual incomes (\$USA)	
Median range	5760
Race/Ethnicity	
Black	64 (98.46%)
White	1(1.53%)
Viral and Bacteria stature	
HIV	
Positive	0%
Négative	100%
HBV	
Positive	0%
Négative	100%
HCV	
Positive	0%
Négative	100%
DENGUE SEROTYPE 1, 2,3 et 4 IgM	
Positive	0%
Négative	100%
Mycobacterium tuberculosis	
Positive	(2/17) 3.07%
Négative	(15/17) 96.93%
Lyme disease	
Positive	0%
Négative	100%
CMV	
Positive	0%
Négative	100%

Table 1: Demographic and clinical characteristics of patients with in the study cohort (n = 65)

fractions of proteins, it's found that the hypoalbuminemia is associated with autoimmune diseases (p=0.02). Thus hypergammaglobulinemia is associated with SLE and hypogammaglobulinemia is associated with Hashimoto Thyroiditis.

Auto antibodies profiles in the sera of patients with autoimmune diseases

The table 3 show the auto antibodies profiles in the sera of patients with autoimmune diseases in the study cohort. Comparing the proportion by type of autoimmune disease, it is found that there is no significant statistical difference in ours cohort (p=0.07).

Association between gender and autoimmune diseases in Burkina Faso

In ours study, there was a predominance of women among the affected member of study. Overall, 94.12% (16/17) of affected individuals were female and 5.88% (1/17) of affected individuals were men. Performances and the diagnosis of Immunodot technique were respectively 100% and 57% for the sensitivity and specificity.

The correct use and interpretation of serologic testing for diagnosing autoimmune diseases present a challenge to clinicians for two reasons: first the sensitivity and specificity of most laboratory tests for autoimmune disease are significantly less than 100% and second the detection of autoantibodies using different techniques such as indirect immune fluorescence or multiplex Immunodot assays give different results.

Autoimmune diseases proportion and patient's socio economic stature

The table 4 show the proportion of autoimmune diseases and patient's socioeconomic status in ours study. Comparing pregnancy's impact on autoimmune disease occurrence, it is found that women who have had multiples pregnancies are more affected than primiparous (p=0.002).

Women's age and autoimmune diseases

The figure 1 show the Women's age and autoimmune diseases.

Comparing different age proportions depending on the autoimmune disease, SLE and SSG were more frequently in young women old of 32.5 years and 35 years respectively (Khi 2=5.30). The old women were more likely to have CREST (68 years), RA (60.4 years), with HT (50 years) and CREST and Polymyositis (46.5 years) (Khi 2 = 3.72).

Discussion

Rate of autoimmune diseases

In this study, the incidence of autoimmune diseases is very low (0.023%) and all except one patient were women. The referral diagnosis was right in 17/65 patients (26.2%) among patients in the limited setting country (Burkina Faso). Data analysis confirmed the low frequency of autoimmune diseases. There were 0.005% patients with systemic lupus erythematosus (SLE), 0.007% with rheumatoid arthritis (RA), 0.003% with CREST and Polymyositis, 0.003% with CREST, 0.001% with Gougerot Sjögren's syndrome (SSG), 0.003% with Thyroiditis of Hashimoto and, 0.001% with sarcoidosis. Autoimmune diseases prevalence is in the range of 4-10% in American and European populations [8,14-18]. Given that the prevalence of autoimmune diseases in USA and Europe is 4-10%, as referred, the 0.023% of autoimmune diseases found is very low and it probably simply indicates that there are many other patients with autoimmune diseases non-diagnosed. We report that SLE and SSG were more frequently in young women old of 32.5 years and 35 years respectively. The old women were more likely to have CREST (68 years), RA (60.4 years), HT (50 years) and CREST and Polymyositis (46.5 years). This gender inequality found in autoimmune diseases is often due to hormonal factors [3-8,19,20];

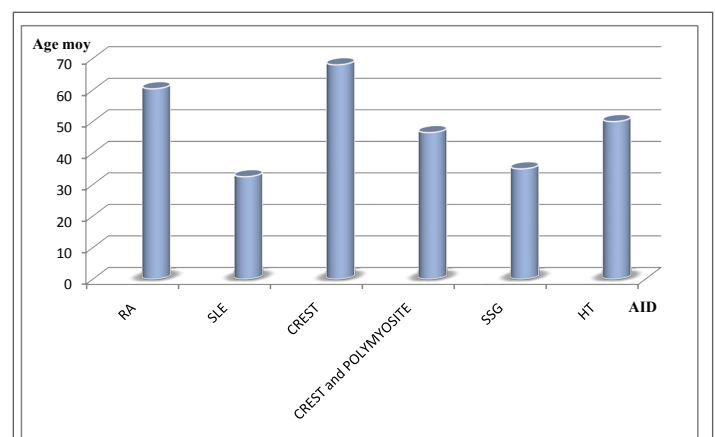


Figure 1: Women's age and autoimmune diseases

Moy. Range of Proteins fractions (g/dl)	RA (n=5)	SLE (n= 4)	SSG (n=1)	Hashimotothyroiditis (n =2)	CREST (n=2)	CREST Sarcoidose and Poly-myositis		References Values
						(n=2)	(n=2)	
Albumin	21.92	22.98	9.62	18.72	17.3	25.75	NA	(40.2-47.6)
Alpha1	2.37	3.06	1.13	2.78	2.62	1.53	NA	(2.1-3.5)
Alpha2	3.4	5.37	1.7	2.88	4.35	3.2	NA	(5.1-8.5)
Beta 1	2.93	1.9	0.79	0.8	2.05	1.89	NA	(3.4-5.2)
Beta 2	3.88	4.28	1.49	1.82	3.2	2.64	NA	(2.3-4.7)
Gamma	13.6	30.4	9.26	4.99	11.48	8.98	NA	(8-13.5)
Proteins totals	49.2	68	24	32	41	44	NA	(60-70)

Table 2: Inflammatory protein profile in the sera of patients with autoimmune diseases

NA: Not Applicable

Moy. Range of Proteins fractions (g/dl)	RA (n=5)	SLE (n= 4)	SSG (n=1)	Hashimotothyroiditis (n =2)	CREST (n=2)	CREST Sarcoidose and Poly-myositis		References Values
						(n=2)	(n=2)	
Albumin	21.92	22.98	9.62	18.72	17.3	25.75	NA	(40.2-47.6)
Alpha1	2.37	3.06	1.13	2.78	2.62	1.53	NA	(2.1-3.5)
Alpha2	3.4	5.37	1.7	2.88	4.35	3.2	NA	(5.1-8.5)
Beta 1	2.93	1.9	0.79	0.8	2.05	1.89	NA	(3.4-5.2)
Beta 2	3.88	4.28	1.49	1.82	3.2	2.64	NA	(2.3-4.7)
Gamma	13.6	30.4	9.26	4.99	11.48	8.98	NA	(8-13.5)
Proteins totals	49.2	68	24	32	41	44	NA	(60-70)

Table 3: Auto antibodies profiles in the sera of patients with autoimmune diseases

Autoimmune disease	Gravidity		Hygienic condition	Frequencies
	Primiparous	Multiparous		
RA		5	No hygiene	(5/73950) 0.007%
SLE	1	4	No hygiene	(4/73950) 0.005 %
SSG		1	No hygiene	(1/73950) 0.001%
CREST		2	No hygiene	(2/73950) 0.003%
CREST and Polymyosite		2	No hygiene	(2/73950) 0.003%
Autoimmune Thyroiditis: Hashimoto		2	No hygiene	(2/73950) 0.003%

Table 4: Rate of autoimmune diseases and patient's socio economic statue

antibodies production and the presence of lymphocytes are higher in women than men [2]. Indeed, in the mouse model of MS, experimental autoimmune encephalomyelitis in mice, disease severity is increased by castration in male mice diffuse and decreased by testosterone implantation in females diffuse [4].

The immunomodulatory effects of gonadal steroids, especially, testosterone, may underlie the female-specific association with autoimmune disorders.

We additionally report positive association with RA, SSG, HT, CREST/ Polymyositis and gravidity. These results were reported in many studies [12,13,21-24]. It suggests that transition from fetal cells through the placenta during pregnancy, and their persistence in the mother after

childbirth can in some cases disturb immunity which explains that the women multiparous are more likely to have autoimmune diseases than those with one pregnancy.

In our study we have found that there is no association with hygiene hypothesis and autoimmune disease. The annual incomes was very low in our study (< 5760\$ USA). Studies that have investigated human autoimmune disease risk in relation to the hygienic degree have found many associations [24].

We also examined a negative association with RA, SLE, CREST/ Polymyositis, HT and infection agents (HIV, HBV, HCV, Lyme disease, Dengue serotype 1, 2, 3 and 4) a positive association with *Mycobacterium tuberculosis* and RA cases (3.07%).

It is a possibility that autoimmune disease and the associated *Mycobacterium tuberculosis* infected may modulate the immune response and RA risk, given some observed correlation between the *Mycobacterium tuberculosis* and autoimmune diseases [1,2,9-13]. It would be of interest to evaluate RA risk in relation to the *Mycobacterium tuberculosis* infection in populations where *Mycobacterium tuberculosis* frequencies are sufficiently high (e.g., Africa tropical); and perhaps more importantly, in any population stratified by gender.

Our findings are, however, consistent with a direct identification of autoantibodies in patients samples coupled with clinical signs. Identification of autoantibodies is essential for the diagnosis of autoimmune diseases in limiting setting country. Clinicians can use these results as guidance for classifying patients and/or for assessing their response to specific therapies, especially in cases where development of targeted biological therapies [12,13,21].

Ours technologies have provided a new approach for autoantibody quantification based on multiplex testing which represents an advantage compared to earlier methods such as line blot assays and conventional one well-one test microtiter ELISA diffuse [3,22,23]. These multiplex assays have made it possible to simultaneously detect multiple biomarkers using a single platform and a single serum sample. In this study we have showed different levels of clinical sensitivity for the detection of the most frequently detected autoantibodies in AID patients.

BMD Test® allows testing of seven antigens in parallel from individual serum samples with high specificity (99.8%). This method provides significant savings in time, avoiding the conventional first screening and post-confirmation algorithm currently being used in most clinical testing laboratories for AID sample testing. BMD requires only 20 µL of sample and follows the same steps as a traditional ELISA assay. Together with low setup costs and a fast sample processing time, BMD provides an affordable alternative to currently available multiplex testing systems. These cost-saving features enable small laboratories with limited budgets to use this technology without a large capital outlay and training of laboratory technicians. BMD have demonstrated an excellent analytical sensitivity (100%) but a bad specificity (57%) and when compared with established ELISA assays [4,8,11]. Moreover, when compared with a line immunoassay, BMD represents the quantitative advantage for clinical management of patients [25-28].

Conclusion

The referral diagnosis was right in 17/65 (26.2%). The incidence was 17/73950 (0.023%) in the limited setting country (Burkina Faso). The very low incidence of auto-immune diseases found is probably due to a number of undiagnosed patients. There was not a significant association between auto-immune diseases and infection; there was not also significant association between auto-immune diseases and hygienic hypothesis.

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Author Contributions

Conceived and designed the experiments: Y Sourabié, Y Traoré, F Fumoux, M S Ouédraogo, and C Sirima

Performed the experiments: Y Sourabié, Y Traoré and M S Ouédraogo

Analyzed the data: Y Sourabié and Y Traoré

Contributed reagents/materials/analysis tools: Y Sourabié, Y Traoré and F Fumoux,

Wrote the paper: Y Sourabié, Y Traoré, F Fumoux, M S Ouédraogo, and C Sirima

Conflict of Interest Statement

We declare that we have no conflict of interest.

References

1. Abreu AS, Fernandes GH, Borba EF, Guedes LKN, Ramos JF, et al. (2014) Musculoskeletal Tuberculosis in Dermatomyositis: Association or Coincidence? *Open J Rheumatol Autoimmune Dis* 4: 58-61.
2. Ahonen P, Miettinen A, Perheentupa J (1987) Adrenal and steroidal cell antibodies in patients with autoimmune polyglandular disease type I and risk of adrenocortical and ovarian failure. *J Clin Endocrinol Metab* 64: 494-500.
3. Engvall E, Perlmann P (1972) Enzyme-linked immunosorbent assay, ELISA. III. Quantitation of specific antibodies by enzyme-linked anti-immunoglobulin in antigen-coated tubes. *J Immunol* 109: 129-135.
4. Forsblad-d'Elia H, Carlsten H, Labrie F, Konttinen YT, Ohlsson C (2009) Low serum levels of sex steroids are associated with disease characteristics in primary Sjögren's syndrome; supplementation with dehydroepiandrosterone restores the concentrations. *J Clin Endocrinol Metab* 94: 2044-2051.
5. Habash-Bseiso DE, Yale SH, Glurich I, Goldberg JW (2005) Serologic testing in connective tissue diseases. *Clin Med Res* 3: 190-193.
6. Haga HJ, Rygh T (1999) The prevalence of hyperprolactinemia in patients with primary Sjögren's syndrome. *J Rheumatol* 26: 1291-1295.
7. Hanly JG, Su L, Farewell V, Fritzler MJ (2010) Comparison between multiplex assays for autoantibody detection in systemic lupus erythematosus. *J Immunol Methods* 358: 75-80.
8. Hanly JG, Thompson K, McCurdy G, Fougere L, Theriault C, et al. (2010) Measurement of autoantibodies using multiplex methodology in patients with systemic lupus erythematosus. *J Immunol Methods* 352: 147-152.
9. Balboni I, Chan SM, Kattah M, Tenenbaum JD, Butte AJ, et al. (2006) Multiplexed protein array platforms for analysis of autoimmune diseases. *Annu Rev Immunol* 24: 391-418.
10. Bossuyt X, Louche C, Wiik A (2008) Standardisation in clinical laboratory medicine: an ethical reflection. *Ann Rheum Dis* 8: 1061-1063.
11. Bossuyt X, Frans J, Hendrickx A, Godefridis G, Westhovens R, et al. (2004) Detection of anti-SSA antibodies by indirect immunofluorescence. *Clin Chem* 12: 2361-2369.
12. Cervera R, Balasch J (2008) Bidirectional effects on autoimmunity and reproduction. *Hum Reprod Update* 14: 359-366.
13. Correa PA, Gomez LM, Cadena J, Anaya JM (2005) Autoimmunity and tuberculosis. Opposite association with TNF polymorphism. *J Rheumatol* 32: 219-224.
14. Forsblad-d'Elia H, Carlsten H, Labrie F, Konttinen YT, Ohlsson C (2009) Low serum levels of sex steroids are associated with disease characteristics in primary Sjögren's syndrome; supplementation with dehydroepiandrosterone restores the concentrations. *J Clin Endocrinol Metab* 94: 2044-2051.
15. Habash-Bseiso DE, Yale SH, Glurich I, Goldberg JW (2005) Serologic testing in connective tissue diseases. *Clin Med Res* 3: 190-193.
16. Haga HJ, Rygh T (1999) The prevalence of hyperprolactinemia in patients with primary Sjögren's syndrome. *J Rheumatol* 26: 1291-1295.
17. Hanly JG, Su L, Farewell V, Fritzler MJ (2010) Comparison between multiplex assays for autoantibody detection in systemic lupus erythematosus. *J Immunol Methods* 358:75-80.

18. Hanly JG, Thompson K, McCurdy G, Fougere L, Theriault C, et al. (2010) Measurement of autoantibodies using multiplex methodology in patients with systemic lupus erythematosus. *J Immunol Methods* 352: 147-152.
19. El Miedany YM, Ahmed I, Moustafa H, El Baddani M (2004) Hyperprolactinemia in Sjögren's syndrome: a patient subset or a disease manifestation? *Joint Bone Spine* 71: 203-208.
20. Endo Y, Negishi I, Ishikawa O (2002) Possible contribution of microchimerism to the pathogenesis of Sjögren's syndrome. *Rheumatology (Oxford)* 41: 490-495.
21. Kumar Y, Bhatia A, Minz RW (2009) Antinuclear antibodies and their detection methods in diagnosis of connective tissue diseases: a journey revisited. *Diagn Pathol* 4: 1.
22. Kumble S, Choi L, Lopez-Muedano C, Kumble KD. (2010) Microarray ELISA for autoantibody screening in connective tissue diseases. *J Clin Diagn Res* 6: 145-149.
23. Okada H, Kuhn C, Feillet H, Bach JF (2010) The 'hygiene hypothesis' for autoimmune and allergic diseases: an update. *Clin Exp Immunol* 160: 1-9.
24. Selmi C, Lu Q, Humble MC (2012) Heritability versus the role of the environment in autoimmunity. *J Autoimmun* 39: 249-252.
25. Fujinami RS, von Herrath MG, Christen U, Whitton JL (2006) Molecular Mimicry, Bystander Activation, or Viral Persistence: Infections and Autoimmune Disease. *Clin Microbiol Rev* 19: 80-94.
26. Vestergaard P (2002) Smoking and thyroid disorders—a metaanalysis. *Eur J Endocrinol* 146: 153–161.
27. Waisberg M, Tarasenko T, Vickers BK, Scott BL, Willcocks LC, et al. (2010) Genetic susceptibility to systemic lupus erythematosus protects against cerebral malaria in mice. *PNAS* 108: 1122-1127.
28. Zafirir Y, Gilburd B, Carrasco MG, Kivity S, Sánchez-Castañón M, et al. (2013) Evaluation of an automated chemiluminescent immunoassay kit for antinuclear antibodies in autoimmune diseases. *Immunol Res* 56: 451-456.